

Syntheses of 1-Deoxynojirimycin-Trehalamine-Fused and -Linked Compounds and Their Biological Activities

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Abstract: 1-Deoxynojirimycin-trehalamine-fused and -linked compounds (10, 19a and 19b) were synthesized from 1-deoxy-2,3,4,6-tetra-O-benzylnojirimycin and trehazolamine, which was obtained from natural trehazolin as a degradation product. None of these synthetic compounds exceeded 1-deoxynojirimycin in the inhibitory activities towards rat intestinal maltase and yeast α -D-glucosidase. © 1998 Elsevier Science Ltd. All rights reserved.

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Introduction

 α -Glucosidases catalyze the regiospecific hydrolysis of α -glucosidic linkages of oligo- and polysaccharides such as starch. Many α -D-glucosidase inhibitors for therapeutic use have been investigated to control diabetes, obesity, HIV, metastasis of cancer, and so on. Practically, acarbose [1,2,3] and AO-128 (voglibose) [4] have been used clinically as drugs controlling hyperglycemia. 1-Deoxynojirimycin was found to be a potent inhibitor of intestinal oligo- and disaccharidases of mammals [5]. And trehazolin, which is a pseudodisaccharide consisting of an α -glucosyl group and a unique aglycon moiety (trehalamine), exhibited powerful inhibitory activity towards various trehalases [6]. We were interested in the structure and D-glycosidase inhibitory activity of both 1-deoxynojirimycin-trehalamine-fused and -linked compounds as

pseudodisaccharides. Here we describe the syntheses of these compounds (10, 19a and 19b) [7] and their biological activities towards rat intestinal maltase.

Results and Discussion

[Syntheses]

Trehazolamine (1), trehazolin aminocyclitol moiety, obtained by hydrolysis of natural trehazolin [8] was treated with benzyl chloroformate in THF-H₂O containing pyridine at 0-5 °C, and the resulted Nbenzyloxycarbonyl compound was converted to the tri-O-silylated compound 2 (11%) and the tetra-O-silylated compound 3 (34%) with tert-butyldimethylsilyl chloride and 4-dimethylaminopyridine (DMAP) in N,Ndimethylfornamide (DMF). Compound 2 was also silylated at 20-25 °C for four days to give 3 in 62% yield accompanying the recovery of 2 (38%). The silvlated position of 2 was determined from the ¹H NMR analysis of 2' after acetylation of the secondary alcohol of 2 by acetic anhydride-pyridine. That is, the C2 proton of 2' appeared at δ 4.98 as a doublet (J=6.2 Hz). This shows that the remaining secondary hydroxy group of 2 was at the C2 position. Hydrogenolysis of 3 using palladium on carbon as a catalyst gave 4 (92%). Treatment of 4 with carbon disulfide, triethylamine and 2-chloro-1-methylpyridinium iodide in dichloromethane gave isothiocyanate 5 (87%) [9] as a solid (mp 47-49 °C) after silica gel chromatography. Reaction of compound 5 with 1-deoxy-2,3,4,6-tetra-O-benzylnojirimycin 6, prepared by the reported method [7], in a small volume of tetrahydrofurane (THF) using triethylamine as a catalyst gave thiourea 7 (68%). Treatment of 7 with tetrabutylammonium fluoride gave pentaol 8 (93%). Treatment of 8 with 2-chloro-3ethylbenzoxazolium tetrafluoroborate and triethylamine in acetonitrile gave a 1-deoxynojirimycin-trehalamine fused oxazoline compound 9 (95%) [10]. Hydrogenolysis of the benzyl groups of 9 using palladium hydroxide on carbon as a catalyst gave an inseparable mixture of 10 and 11 after chromatography using Amberlite CG-50 (NH₄+ type/H+ type = 3/2) followed by lyophilization.

In the ^{1}H NMR, the spectral peak intensity of 10 gradually increased during 4 days, while that of 11 gradually decreased, and the mixture of 10 and 11 in an NMR tube (D₂O) was initially 1: 4, and finally reached an equilibrium at the ratio 4: 1. ^{1}H - ^{13}C long range couplings were observed between C2-carbon (169.8 ppm) and 3a, 6a-two protons of compound 10. In the ^{1}H - and ^{13}C -NMR studies, it is coincident with the structure of 11 that the chemical shifts of C6-two protons and C6-carbon of 1-deoxynojirimycin moiety of 11 are shifted to lower fields than those of 10. That is, the chemical shifts of the C6-protons of 10 are δ 3.62 and 3.80, and those of 11 are δ 4.30 and 4.44. The chemical shift of the C6-carbon of 10 is δ 61.7 and that

Scheme 1

Reagents and conditions: R = tert-BuMe₂Si; a) ClCOOBn, pyridine, H_2O -THF (2:1), 0-5 °C, 30 min, concentrated; then tert-BuMe₂SiCl, DMAP, DMF, 20-25 °C, 16 h, **2**, 34%, **3**, 11%; b) tert-BuMe₂SiCl, DMAP, DMF, 20-25 °C, 4 days, ca. 62% (recovery **2**, 38%); c) H_2 , Pd/C, THF, 24 °C, 8 h, quant.; d) CS_2 , Et_3N , 2-chloro-1-methylpyridinium iodide, CH_2Cl_2 , 24 °C, 2.5 h, 87%; e) Et_3N , THF, 60-65 °C, 3 h, 68% (recovery of **5**, 22% and **6**, 19%); f) n-Bu₄NF, THF, 24 °C, 3 h, 93%; g) 2-chloro-3-ethylbenzoxazolium tetrafluoroborate, Et_3N , MeCN, 0 °C, 10 min, 95%; h) H_2 , $Pd(OH)_2/C$, MeOH, 60 °C, 40 min, an inseparable 4:1 equilibrium mixture of **10** and **11**, 32%.

Scheme 2

Reagents and conditions: R = tert-BuMe₂Si; a) N-Boc-glycine or N-Boc- β -alanine, DCC, DMAP, CH_2Cl_2 , 24 °C, 6 h, 12a, 100%, 12b, 98%; b) BH_3 -THF complex, THF, 24 °C, 16 h, 13a, 62%, 13b, 98%; c) CF_3COOH , CH_2Cl_2 , 24 °C, 45 min, 14b, 95%; d) CS_2 , Et_3N , CH_2Cl_2 , 24 °C, 1 h, then Et_3N , 2-chloro-1-methylpyridinium iodide, two steps 41%; e) 4, cat. Et_3N , THF, 20-25 °C, 2 days, 87%; f) THF, 60-65 °C, 3 h, 83%; g) HCl-MeOH, 24 °C, 16 h, quant.; h) 2-chloro-3-ethylbenzoxazolium tetrafluoroborate, Et_3N , MeCN, 0 °C, 30 min, 18a, 67%, 18b, 77%; i) H_2 , $Pd(OH)_2/C$, MeOH, 60-65 °C, 8 h, 19a, 72%, 50-55 °C, 24 h, 19b, 98%.

of 11 is δ 70.7. And also the chemical shift of the C6a-carbon (90.2 ppm) of the trehalamine part of 10 is shifted to a lower field than that of 11 (88.1 ppm). This type of 2-amino-2-oxazoline ring migration reaction from 10 to 11 should be possible according to our previous studies [11,12].

The compounds 19a and 19b were synthesized as follows. Condensation of 6 with *N-tert*-butoxycarbonylglycine or *N-tert*-butoxycarbonyl-β-alanine using 1,3-dicyclohexylcarbodiimide (DCC) and DMAP gave 12a or 12b in good yields. Reduction of amide-carbonyl of 12a or 12b was performed using BH3-THF complex to yield amine 13a or 13b. Deprotection of *N-tert*-butoxycarbonyl group from 13a or 13b was accomplished by treatment with trifluoroacetic acid to give 14a or 14b. The amine part of 14a was converted to an isothiocyanate 15a by treatment with CS2, Et3N, and 2-chloro-3-ethylbenzoxazolium tetrafluoroborate. Reaction of 15a with amine 4 in THF at 20-25 °C for 2 days using Et3N as a catalyst gave thiourea 16a. On the other hand, reaction of amine 14a with isothiocyanate 5 in THF at 60-65 °C gave thiourea 16b. Deprotection of *tert*-butyldimethylsilyl groups on trehazolamine moiety of 16a and 16b by HCl-MeOH gave tetraols 17a and 17b, respectively. Oxazoline formation of 17a and 17b by 2-chloro-3-ethylbenzoxazolium tetrafluoroborate and Et3N in MeCN yielded 18a and 18b, respectively. Hydrogenolysis of benzyl groups of 18a and 18b gave 19a and 19b, respectively.

N-Aminoalkyl derivative of 1-deoxynojirimycin (20) and N-alkyltrehalamine (23), which are lacking respective counterparts, were also prepared to compare the biological activities of these compounds with that of 1-deoxynojirimycin. Hydrogenolysis of benzyl group of 14b yielded 20, and the procedure of preparation of 23 was as follows. Reaction of the amine 4 with butyl isothiocyanate in THF using Et₃N as a catalyst gave thiourea 21 (mp 141-142 °C). Subsequent desilylation of 21 with HCl-MeOH gave tetraol 22. Treatment of 22 with 2-chloro-3-ethylbenzoxazolium tetrafluoroborate and Et₃N in MeCN yielded 23.

Scheme 3

Reagents and conditions: R = tert-BuMe₂Si; a) H_2 , $Pd(OH)_2/C$, MeOH, 50-55 °C, 16 h, 86%, b) n-C₄ H_9 N=C=S, Et_3 N, THF, rt, 16 h, 73%; c) HCl-MeOH, 24 °C, 4 h; d) 2-chloro-3-ethylbenzoxazolium tetrafluoroborate, Et_3 N, MeCN, 0 °C, 30 min, two steps 46%.

[Biological Activities]

Inhibition of glycosidases (rat intestinal maltase and yeast α -D-glucosidase) by synthetic compounds (a mixture of 10 and 11, 19a, 19b, 1-deoxynojirimycin, 20, and 23) was assayed.

For the measurement of rat intestinal maltase, a glucose oxidase-peroxidase system [13,14] was applied. For dilution, 20 mM phosphate buffer (pH 6.2) was used, and the reactions for assay were carried out at 37 °C for 20 min in a 96-well microtiter plate in a total volume of 150 µL, composed from 60 µL of phosphate buffer, 10 µL of varying concentrations of inhibitors, 20 µL of 39.5 mM maltose, 20 µL of 1 x 10⁻³ units of rat intestinal maltase (added last), and glucose assay solution (diluted with 40 µL of phosphate buffer) containing 3 units of glucose oxidase (Boehringer Mannheim), 0.13 units of horseradish peroxidase (Boehringer Mannheim), 40 µg of phenol and 20 µg of 4-aminoantipyrine (Sigma). Then, colorimetric changes at 492 nm were measured by micoplate reader, and the amount of glucose released was calculated. The enzyme inhibitory activities of a mixture of 10 and 11, 19a, 19b, 20, 23, and 1-deoxynojirimycin exhibited IC50 values of 0.68, 4.2, 1.5, 1.4, 9.1, and 0.023 µg/mL, respectively.

For the measurement of yeast α -glucosidase inhibition, 20 mM phosphate buffer (pH 6.2) was used to dilute the reagents, and the reactions for assay were carried out at 37 °C for 10 min in a 96-well microtiter plate in a total volume of 150 μ L composing from 70 μ L of phosphate buffer, 10 μ L of varying concentrations of inhibitors, 50 μ L of 10 mM 4-nitrophenyl α -D-glucopyranoside, and 20 μ L of yeast α -D-glucosidase (0.03 U/mL, added finally). Then, 20 μ L of 4M sodium glycinate buffer (pH 10.4) was added to stop the reactions, and colorimetric changes at 415 nm being directly proportional to the amount of liberated 4-nitrophenol were measured by micoplate reader. The enzyme inhibitory activities of a mixture of 10 and 11, 19a, 19b, 20, 23, and 1-deoxynojirimycin exhibited IC50 values of >200, >200, >200, >200, 42, and 13 μ g/mL, respectively.

Conclusion

We have synthesized the 1-deoxynojirimycin-trehalamine-fused and -linked compounds 10, 19a and 19b, and also the N-aminoalkyl derivative of 1-deoxynojirimycin (20) and N-alkyltrehalamine (23), which are lacking respective counterparts. And the biological activities of these compounds were compared with those of 1-deoxynojirimycin. None of them exceeded 1-deoxynojirimycin in the inhibitory activities towards rat intestinal maltase and yeast α -D-glucosidase.

Experimental

Melting points were determined on a Yanagimoto micro melting point apparatus and are uncorrected. ¹H NMR (270, 400 and 500 MHz) spectra were recorded with JEOL JNM-270, JEOL JNM-GSX 400 and JEOL GSX 500 spectrometers using TMS as an internal standard. IR absorption spectra were determined with a IR A-2 spectrophotometer, and mass spectra were obtained with a JMS-700 mass spectrometer. Optical rotations were obtained by the use of a Perkin-Elmer 241 polarimeter. Elemental analyses were performed by the

Institute of Science and Technology, Inc. Separation of the compounds by column chromatography was done with silica gel 60 (230-400 mesh ASTM, E. Merck) under a slightly elevated pressure (1.2-1.5 atm) for easy elution, and the quantity of silica gel used was 50-100 times of the weight charged on the column. Preparative TLC was performed on silica gel plates (Merck, Silica Gel 60 F₂₄₅). Detection involved spraying the chromatogram with a solution of 17% H₂SO₄ in water (w/w), containing ammonium molybdate (2.3%) and ceric sulfate (0.9%) (Hanessian dip), and heating the plate for several minutes at ca 180 °C. Tetrahydrofurane (THF) was distilled from LiAlH₄ and used immediately. Dichloromethane was dried by being passed through an ICN Alumina B-Super I. N,N-Dimethylformamide (DMF) and pyridine were dried by storage over 4Å molecular sieves. Acetonitrile was dried by storage over 3Å molecular sieves.

 $[1R-(1\alpha,2\beta,3\alpha,4\beta,5\beta)]$ -5-tert-Butoxycarbonylamino-1-(tert-butyldimethylsilyloxymethyl)-3,4-bis(tert-butyldimethylsilyloxy)cyclopentan-1,2-diol [1R - $(1\alpha,2\beta,3\alpha,4\beta,5\beta)$]-5-tert-Butoxycarbonylamino-1-(tert-butyldimethylsilyloxymethyl)-2,3,4-To a solution of trehazolamine (1) [8] tris(tert-butyldimethylsilyloxy)cyclopentan-1-ol (3). (3.60 g, 20.1 mmol) in H₂O-THF (2:1, 165 ml) and pyridine (14 g, 177 mmol) was added ClCOOBn (11.0 g, 64.5 mmol) at 0-5 °C with vigorous stirring. The stirring was continued for 30 min at this temperature, then the mixture was concentrated in vacuo, and dried with a high vacuum pump. The residue was dissolved in DMF (20 ml), and to this solution were added DMAP (18.0 g, 147 mmol) and t-BuMe₂SiCl (18.0 g, 119 mmol). The mixture was stirred for 16 h at 20-25 °C, diluted with excess EtOAc, washed sequentially with aqueous 0.5M HCl, H2O, sat. NaHCO3, and brine, dried over MgSO4, filtered, and concentrated in vacuo to give a mixture, which was chromatographed on a silica gel column. Elution with cyclohexane-EtOAc (10:1) gave 3 (5.29 g, ca 34% yield, containing a small amount of t-BuMe₂SiOBn), and further elution with cyclohexane-EtOAc (4:1) gave 2 (1.40 g, 11% yield). Physical data of 3: IR v_{max} (film) 3450, 3400, 1725, 1708 cm⁻¹. 270 MHz ¹H NMR (CDCl₃) δ 0.06-0.12 (24H, m), 0.88 (9H, s), 0.89 (9H, s), 0.90 (18H, s), 3.06 (1H, s, OH), 3.74-3.79 (2H, m), 3.93 (1H, d, J=2.0 Hz), 4.08 (1H, s), 4.12 (2H, s), 5.06, 5.12 (2H, AB-q, J=12.5 Hz), 6.10 (1H, d, J=7.2 Hz, NH), 7.22-7.40 (5H, m). FAB MS (positive); m/z 770 [M+H]⁺. Anal. Calcd. for C38H75NO7Si4 (770.4): C, 59.25; H, 9.81; N, 1.82. Found: C, 58.97; H, 9.60; N, 1.86. Physical data of 2: IR ν_{max} (film) 3420, 2950, 2920, 2880, 2860, 1700 cm⁻¹. ¹H NMR (CDCl₃) δ 0.06-0.11 (18H, m), 0.87 (9H, s), 0.88 (18H, s), 2.78 (1H, d, J=4.3 Hz, OH), 3.24 (1H, s, OH), 3.79-3.92 (4H, m), 4.03-4.09 (2H, m), 5.09 (2H, s), 7.32 (5H, bs). FAB MS (positive); m/z 656 [M+H]+. Anal. Calcd. for C₃₂H₆₁NO₇Si₃ (656.1): C, 58.58; H, 9.37; N, 2.14. Found: C, 58.24; H, 9.16; N, 2.11.

[1R- $(1\alpha,2\beta,3\alpha,4\beta,5\beta)$]-5-tert-Butoxycarbonylamino-1-(tert-butyldimethylsilyloxymethyl)-2,3,4-tris(tert-butyldimethylsilyloxy)cyclopentan-1-ol (3).

To a solution of 2 (926 mg, 1.41 mmol) in DMF (8 ml) were added DMAP (259 mg, 2.11 mmol) and t-BuMe₂SiCl (320 mg, 2.11 mmol). The mixture was stirred for 4 days at 20-25 °C, diluted with excess EtOAc, washed with aqueous 0.5M HCl, H₂O, sat. NaHCO₃, and brine, dried over MgSO₄, filtered, and concentrated *in vacuo* to give a residue, which was chromatographed on a silica gel column. Elution with cyclohexane-EtOAc (10:1) gave 3 (679 mg, ca. 62%), which contained a small amount of benzyl tert-

butyldimethylsilyl ether. This mixture was also employed for the next reaction without further purification. Recovery of the starting 2 was 356 mg (38%).

 $[1R-(1\alpha,2\beta,3\alpha,4\beta,5\beta)]$ -2-Acetoxy-5-tert-butoxycarbonylamino-1-(tert-

butyldimethylsilyloxymethyl)-3,4-bis(tert-butyldimethylsilyloxy)cyclopentane-1-ol (2') A solution of 2 (10 mg) in pyridine (0.4 ml) and acetic anhydride (0.2 ml) was allowed to stand overnight at room temperature. The reaction mixture was concentrated in vacuo to give a residue, which was chromatographed on a silica gel small column. Elution with cyclohexane-EtOAc (4:1) gave 2' (8 mg, 75%) as an oil. IR v_{max} (film) 3448, 2950, 2925, 2885, 2860, 1727 cm⁻¹. 270 MHz ¹H NMR (CDCl₃) δ 0.03 (3H, s), 0.04 (3H, s), 0.08 (3H, s), 0.10 (6H, s), 0.91 (9H, s), 0.92 (9H, s), 0.93 (9H, s), 2.14 (3H, s), 3.59, 3.75 (2H, AB-q, J=10.2 Hz), 4.08-4.18 (2H, m), 4.30 (1H, dd, J=7.7, 8.1 Hz), 4.98 (1H, d, J=6.2 Hz, C2-H), 5.14 (2H, s), 5.68 (1H, d, J=8.1 Hz), 7.36 (5H, bs). FAB MS (positive); m/z 720 [M+Na]+, 698 [M+H]+. Anal. Calcd. for C34H63NO8Si3 (698.1): C, 58.50; H, 9.10; N, 2.01. Found: C, 58.55; H, 9.15; N, 2.05.

[1R-(1 α ,2 β ,3 α ,4 β ,5 β)]-5-Amino-1-(*tert*-butyldimethylsilyloxymethyl)-2,3,4-tris(*tert*-butyldimethylsilyloxy)cyclopentan-1-ol (4). A solution of 3 (1.36 g, 1.77 mmol) in THF (100 ml) containing 10% Pd on carbon (0.7 g) as catalyst was stirred for 8 h at 24 °C under an atmosphere of hydrogen, and filtered, concentrated *in vacuo* to give 4 (1.12 g, quantitative) as a solid. mp 79-82 °C. IR ν_{max} (Nujol) 3560 (w), 3400 (w) cm⁻¹. 270 MHz ¹H NMR (CDCl₃) δ 0.09-0.11 (24H, m), 0.89 (9H, s), 0.90 (9H, s), 0.91 (9H, s), 0.93 (9H, s), 1.66 (2H, bs, NH₂), 3.00 (1H, bs, OH), 3.14 (1H, d, J=5.2 Hz, C5-H), 3.76, 3.99 (2H, AB-q, J=10.3 Hz), 3.83 (1H, d, J=3.4 Hz, C2-H), 3.90 (1H, dd, J=3.4, 3.8 Hz), 4.00 (1H, dd, J=3.8, 5.2 Hz). FAB MS (positive); m/z 636 [M+H]⁺. *Anal.* Calcd. for C₃₀H₆₉NO₅Si₄ (636.2): C, 56.64; H, 10.93; N. 2.20. Found: C, 56.60; H, 10.79; N, 2.14.

[1R-(1 α ,2 β ,3 α ,4 β ,5 β)]-1-(tert-Butyldimethylsilyloxymethyl)-5-isothiocyanato-2,3,4-tris(tert-butyldimethylsilyloxy)cyclopentan-1-ol (5). To a solution of 4 (508 mg, 0.798 mmol) in CH₂Cl₂ (20 ml) were added CS₂ (122 mg, 1.60 mmol) and Et₃N (202 mg, 2.00 mmol) at 24 °C under nitrogen. The solution was stirred for 1 h, and then to this solution were added Et₃N (162 mg, 1.60 mmol) and 2-chloro-1-methylpyridinium iodide (307 mg, 1.20 mmol). This solution was stirred for 2.5 h at 24 °C under nitrogen, concentrated in vacuo, diluted with EtOAc, washed with sat. NaHCO₃, H₂O, brine, dried over MgSO₄, filtered, and concentrated in vacuo to give a mixture, which was chromatographed on a silica gel column. Elution with cyclohexane-EtOAc (24:1) gave 5 (473 mg, 87%). IR v_{max} (film) 3520 (w), 2960, 2935, 2900, 2860, 2125 cm⁻¹. 270 MHz ¹H NMR (CDCl₃) δ 0.09-0.11 (24H, m), 0.89 (9H, s), 0.91 (9H, s), 0.93 (9H, s), 0.95 (9H, s), 3.44 (1H, bs, OH), 3.73, 3.95 (2H, AB-q, J=10.3 Hz), 3.83-3.90 (2H, m), 3.96 (1H, d, J=5.6 Hz), 4.10 (1H, dt, J=<1, 4.0 Hz). FAB MS (positive); m/z 678 [M+H]⁺, 620. Anal. Calcd. for C₃₁H₆₇NO₅SSi₄ (678.3): C, 54.89; H, 9.96; N, 2.07; S, 4.73. Found: C, 54.59; H, 9.69; N, 2.09; S, 4.51.

N-[[1R-(1 α ,2 β ,3 α ,4 β ,5 β)]-[1-(tert-Butyldimethylsilyloxymethyl)-1-hydroxy-2,3,4-tris(tert-butyldimethylsilyloxy)cyclopent-5-yl]aminothiocarbonyl]-2,3,4,6-tetra-O-benzyl-1,5-dideoxy-1,5-imino-D-glucitol (7). A THF solution of 5 (52.4 mg, 0.100 mmol) and 2,3,4,6-

tetra-O-benzyl-1-deoxynojirimycin (6, 67.8 mg, 0.100 mmol) was concentrated *in vacuo*. To the residue were added THF (0.10 ml) and Et₃N (9.5 mg). The mixture was maintained at 60-65 °C for 3 h with stirring, and after evaporation, the residue was chromatographed on a silica gel TLC plate. Development with cyclohexane-EtOAc (10:1) gave 7 (82 mg, 68%) at R_f=0.41, starting 5 (15 mg, 22% recovery) and tetra-O-benzyl-1-deoxynojirinycin (10 mg, 9% recovery). IR v_{max} (film) 3365, 2955, 2930, 2888, 2859 cm⁻¹. 270 MHz ¹H NMR (CDCl₃) δ 0.09 (3H, s), 0.12 (3H, s), 0.15 (3H, s), 0.17-0.19 (12H, m), 0.24 (3H, s), 0.88 (9H, s), 0.92 (9H, s), 0.96 (9H, s), 1.00 (9H, s), 3.16 (1H, s, OH), 3.59-3.75 (3H, m), 3.81-3.93 (3H, m), 3.96-4.07 (2H, m), 4.10 (1H, s), 4.21 (1H, m), 4.38-4.85 (11H, m), 4.98 (1H, m), 7.11 (1H, d, J=6.6 Hz, NH), 7.34 (20H, s). FAB MS (positive); m/z 1202, 1201 [M+H]⁺. Anal. Calcd. for C₆₅H₁₀₄N₂O₉SSi₄ (1202.0): C, 64.95; H, 8.72; N, 2.33; S, 2.67. Found: C, 65.28; H, 8.75; N, 2.34; S, 2.63.

N-[[1R-(1 α ,2 β ,3 α ,4 β ,5 β)]-[1-Hydroxymethyl-1,2,3,4-(tetrahydroxy)cyclopent-5-yl]aminothiocarbonyl]-2,3,4,6-tetra-O-benzyl-1,5-dideoxy-1,5-imino-D-glucitol (8). To a solution of 7 (148 mg, 0.123 mmol) in THF (6 ml) was added 1M THF solution of n-Bu₄NF (1.20 ml, 1.20 mmol). The solution was stirred for 3 h at 24 °C, diluted with EtOAc, washed with brine, dried over MgSO₄, and filtered, and the filtrate was concentrated *in vacuo* to give a residue, which was chromatographed on a silica gel column. Elution with EtOAc, and then 5% MeOH in EtOAc gave 8 (85 mg, 93%). IR ν_{max} (film) 3360, 2924, 2870 cm⁻¹. 270 MHz ¹H NMR (CDCl₃) δ 3.46-3.80 (10H, m), 3.96-4.04 (3H, m), 4.31-4.73 (8H, m), 4.84 (1H, t, J=6.2 Hz, changed to a doublet on addition of D₂O), 7.15-7.31 (21H, m, containing NH). FAB MS (positive); m/z 745 [M+H]⁺. Anal. Calcd. for C₄₁H₄₈N₂O₉S (744.9): C, 66.11; H, 6.94; N, 3.76; S, 4.30. Found: C, 66.25; H, 6.87; N, 3.77; S, 4.12.

N-[[3aR-(3a α ,4 α ,5 β ,6 α ,6a α)]-4-Hydroxymethyl-3a,5,6,6a-tetrahydro-4,5,6-trihydroxy-4H-cyclopentoxazol-2-yl]-2,3,4,6-tetra-O-benzyl-1,5-dideoxy-1,5-imino-D-glucitol (9).

To a solution of **8** (52 mg, 0.070 mmol) in MeCN (3.5 ml) was added 2-chloro-3-ethylbenzoxazolium tetrafluoroborate (34.7 mg, 0.129 mmol) at 0 °C. After stirring for 1 h at 0 °C, Et₃N (35 mg, 0.347 mmol) was added to this solution at 0 °C. After stirring for 10 min at 0 °C, the reaction mixture was concentrated *in vacuo*, and chromatographed on a silica gel column. Elution with 10% MeOH in EtOAc gave 9 (47 mg, 95%). IR v_{max} (film) 3280 (broad), 1641 cm⁻¹. 270 MHz ¹H NMR (CDCl₃+DMF-d₇+D₂O) δ 3.62-4.83 (22H, m), 7.21-7.32 (20H, m). FAB MS (positive); m/z 733 [M+Na]⁺, 711 [M+H]⁺. Anal. Calcd. for C₄₁H₄₆N₂O₉ (710.8): C, 69.28; H, 6.52; N, 3.94. Found: C, 69.17; H, 6.31; N, 3.90.

N-[[3aR-(3a α ,4 α ,5 β ,6 α ,6a α)]-4-Hydroxymethyl-3a,5,6,6a-tetrahydro-4,5,6-trihydroxy-4H-cyclopentoxazol-2-yl]-1,5-dideoxy-1,5-imino-D-glucitol (10) and N,6-O-[(trehazolaminyl)iminomethylidene]-1-deoxynojirimycin (11). A solution of 9 (50 mg, 0.070 mmol) in MeOH (10 ml) containing Pd(OH)₂ on carbon (1.0 g, wet, Degussa type, Pd content 20%) was stirred for 40 min at 60 °C under an atmosphere of hydrogen. The mixture was filtered. The catalyst was washed with MeOH (5 ml). The combined filtrate was concentrated *in vacuo*. The residue was chromatographed on Amberlite CG 50 (NH₄+ type/H+ type = 3/2, 20 ml). Elution with 0.5 M NH₃ aqueous solution (4 ml each) gave an inseparable mixture of 10 and 11 (8.0 mg, 32%) as a powder after lyophilization. The mixture of 10 and 11 in an NMR tube (D₂O) equilibrated at 4:1 after 4 days. IR v_{max} (Nujol) 3370,

1665 (shoulder), 1640 cm-1. FAB MS (positive); m/z 351. High resolution FAB MS (positive); Calcd. for C₁₃H₂₃N₂O₉: 351.1403. Found: 351.1383. 500 MHz ¹H and ¹³C NMR data of 10: trehalamine part; ¹H NMR (D₂O) δ 3.77 (1H, m, C4CHHOH), 3.87 (1H, m, C4CHHOH), 3.96 (1H, m, C5H), 4.25 (1H, m, C6H), 4.25 (1H, d, $J_{3a.6a}$ =8.9 Hz, C3aH), 5.00 (1H, d, $J_{3a.6a}$ =8.9 Hz, C6aH); 1-deoxynojirimycin part; δ 2.44 (1H, d, $J_{geminal}$ =11.4 Hz, C1HHN), 2.52 (1H, m, C5H), 3.10 (1H, dd, J=4.7, 11.4 Hz, C1HHN), 3.22 (1H, $J_{3,4}$ =9.5 Hz, C4H), 3.32 (1H, t, J=9.5 Hz, C3H), 3.45 (1H, dd, J=4.7, 9.5 Hz, C2H), 3.62 (1H, m, C6HHOH), 3.80 (1H, m, C6HHOH); trehalamine part; ¹³C NMR (D₂O) δ 62.3 (C4CH₂OH), 65.6 (C3a), 82.8 (C5), 83.2 (C6), 85.4 (C4), 90.2 (C6a); 1-deoxynojirimycin part: δ 51.7 (C1), 61.7 (C6), 63.7 (C5), 70.5 (C2), 74.2 (C4), 81.8 (C3). 500 MHz ¹H and ¹³C NMR data of 11: trehazolamine part: ¹H NMR (D₂O) δ 3.58 (1H, m, CHHOH), 3.75 (1H, m, CHHOH), 3.90 (1H, m, C3H), 4.17 (1H, m, C2H), 4.33 (1H, m, C5H), 4.92 (1H, m, C4H); 1-deoxynojirimycin part; δ 2.84 (1H, m, C1HHN), 3.38 (1H, m, C4H), 3.42 (1H, m, C3H), 3.53 (1H, m, C5H), 3.63 (1H, m, C5H), 3.87 (1H, m, C1HHN), 4.30 (1H, m, C6H), 4.44 (1H, m, C6H); trehazolamine part; ¹³C NMR (D₂O) δ 61.8 (C1CH₂OH), 77.0 (C5-N), 83.0 (C3), 83.2 (C2), 88.1 (C4), 80-90 (C1); 1-deoxynojirimycin part: δ 48.3 (C1), 70.7 (C6), 72.0 (C2), 74.5 (C4), 76.6 (C5), 79.8 (C3). In this 500 MHz NMR, the partial structures, determined by the ¹H-¹H connectivity, were elucidated by the double quantum filtered COSY and total correction spectroscopy. The assignment of 13C signals directly bonded to those protons was determined by the heteronuclear multiple-quantum correlation spectrum.

N-[N-(tert-Butoxycarbonyl)glycyl]-2,3,4,6-tetra-O-benzyl-1,5-dideoxy-1,5-imino-D-glucitol (12a). To a solution of 6 (524 mg, 1.00 mmol) in CH₂Cl₂ (10 ml) were added N-(tert-butoxycarbonyl)glycine (210 mg, 1.20 mmol), and DCC (248 mg, 1.20 mmol). The mixture was stirred for 16 h at 24 °C, filtered, and the filtrate was diluted with EtOAc. The solution was washed with 1M aqueous HCl solution, H₂O, sat. aqueous NaHCO₃, and brine, died over MgSO₄, and filtered. The filtrate was concentrated *in vacuo*, and chromatographed on a silica gel column. Elution with cyclohexane-EtOAc (3:1) gave 12a (727 mg, containing a small amount of 1,3-dicyclohexylurea) as a gum, which was employed for the next reaction without further purification. IR v_{max} (film) 3425, 3340 (broad), 1711, 1651 cm⁻¹. 270 MHz ¹H NMR (CDCl₃) δ 1.45 (9H, s), 3.37-3.74 (6H. m), 3.86-4.19 (2H, m), 4.29-4.76 (10H, m), 5.52 (1H, bs, NH), 7.21-7.35 (20H, m). FAB MS (positive); m/z 681 [M+H]⁺, 703 [M+Na]⁺. High resolution FAB MS (positive); m/z Calcd. for C4₁H₄9N₂O₇: 681.3540. Found: 681.3574.

N-[3-(tert-Butoxycarbonylamino)propionyl]-2,3,4,6-tetra-O-benzyl-1,5-dideoxy-1,5-imino-D-glucitol (12b). To a solution of 6 (524 mg, 1.00 mmol) in CH₂Cl₂ (10 ml) were added N-(tert-butoxycarbonyl)-β-alanine (227 mg, 1.20 mmol), DMAP (147 mg, 1.20 mmol) and DCC (248 mg, 1.20 mmol). The mixture was stirred for 6 h at 24 °C, filtered, concentrated in vacuo, and chromatographed on a silica gel column. Elution with cyclohexane-EtOAc (3:2) gave 12b (683 mg, 98%) as a gum. IR ν_{max} (film) 3584-2869, 1711, 1642 cm⁻¹. 270 MHz ¹H NMR (CDCl₃) δ 1.42 (9H, s), 1.75 (1H, broad, NH), 2.53 (2H, bs), 3.32-4.12 (8H, m), 4.25-4.80 (9H, m), 5.31-5.38 (1H, m), 7.21-7.34 (20H, m). Anal. Calcd. for C₄₂H₅₀N₂O₇: C, 72.60; H, 7.25; N, 4.03. Found: C, 72.13; H, 7.31; N, 4.07.

N-[2-(tert-Butoxycarbonylamino)ethyl]-2,3,4,6-tetra-O-benzyl-1,5-dideoxy-1,5-imino-D-glucitol (13a). A solution of the above obtained 12a (727 mg) in 1M BH3-THF complex (30 ml) was stirred for 16 h at 24 °C, and diluted with EtOAc (400 ml). This solution was washed with 1M aqueous HCl (43 ml) by vigorous stirring, and then neutralized with sat. NaHCO3 (60 ml). The organic layer was dried over MgSO4, filtered, concentrated in vacuo, and chromatographed on a silica gel column. Elution with cyclohexane-EtOAc (3:1) gave 13a (412 mg, 62% for two steps, containing a small amount of 1,3-dicyclohexylurea) as a gum. IR v_{max} (film) 3360 (broad), 1711, 1655 cm⁻¹. 270 MHz ¹H NMR (CDCl₃) δ 1.43 (9H, s), 2.17 (1H, t, J=10.9 Hz), 2.40 (1H, d, J=8.9 Hz), 2.47 (1H, td, J=4.9, 13.3 Hz), 2.81-2.91 (1H, m), 3.05-3.25 (3H, m), 3.45-3.66 (5H, m), 4.33-4.98 (8H, m), 5.13 (1H, bs, NH), 7.09-7.13 (2H, m), 7.24-7.36 (18H, m). FAB MS (positive); m/z 667 [M+H]+. High resolution FAB MS (positive); Calcd. for C41H51N2O6: 667.3744. Found: 667.3754. Anal. Calcd. for C41H50N2O6 (666.9): C, 73.85; H, 7.56; N, 4.20. Found: C, 73.46; H, 7.55; N, 4.15.

N-[3-(tert-Butoxycarbonylamino)propyl]-2,3,4,6-tetra-O-benzyl-1,5-dideoxy-1,5-imino-D-glucitol (13b). To compound 12b (695 mg, 1.00 mmol) was added 1M BH3-THF complex (40 ml). The solution was treated as described above to give 13b (683 mg, 98%) as a gum. IR v_{max} (film) 3384-2809, 1711 cm⁻¹. 270 MHz ¹H NMR (CDCl₃) δ 1.25 (2H, bs), 1.43 (9H, s), 2.02 (1H, t, J=10.4-11.0 Hz), 2.24 (1H, d, J=9.1 Hz), 2.37 (1H, m), 2.81 (1H, m), 2.95-3.07 (2H, m), 3.16 (1H, m), 3.43-3.67 (5H, m), 4.34, 4.96 (2H, AB-q, J=10.9 Hz), 4.45, 4.54 (2H, AB-q, J=12.1 Hz), 4.63, 4.70 (2H, AB-q, J=11.6 Hz), 4.82 (2H, t, J=11.5 Hz), 5.25 (1H, bs, NH), 7.08-7.13 (2H, m), 7.26-7.34 (18H, m). FAB MS (positive); m/z 681 [M+H]⁺. High resolution FAB MS (positive); Calcd. for C42H53N2O6: 681.3904. Found: 681.3872. *Anal.* Calcd. for C42H52N2O6 (680.9): C, 74.09; H, 7.70; N, 4.11. Found: C, 74.01; H, 7.85; N, 4.10.

N-(2-Aminoethyl)-2,3,4,6-tetra-O-benzyl-1,5-dideoxy-1,5-imino-D-glucitol (14a). To a solution of the above-obtained 13a (412 mg) in CH₂Cl₂ (7 ml) was added CF₃COOH (3.5 ml). The solution was stirred for 30 min at 24 °C, and diluted with Et₂O (100 ml), washed with 1 M aqueous NaOH, and dried over MgSO₄, filtered, concentrated *in vacuo* to give crude 14a (412 mg) as a powder, which was employed for the next reaction without further purification.

N-(3-Aminopropyl)-2,3,4,6-tetra-O-benzyl-1,5-dideoxy-1,5-imino-D-glucitol (14b). To a solution of 13b (360 mg, 0.529 mmol) in CH₂Cl₂ (15 ml) was added CF₃COOH (3 ml). After the solution was stirred for 45 min at 24 °C, the reaction mixture was treated above to give 14b (291 mg, 95%) as a gum. IR v_{max} (film) 3600-2780, 1715-1580 (w, broad) cm⁻¹. 270 MHz ¹H NMR (CDCl₃) δ 1.53 (1H, m), 1.43 (9H, s), 1.63 (1H, m), 1.85 (2H, bs, NH₂, D₂O exchanged), 2.14 (1H, t, J=10.6 Hz), 2.27 (1H, d, J=9.2 Hz), 2.51 (1H, ddd, J=4.6, 8.6, 13.2 Hz), 2.63 (2H, t, J=6.6 Hz), 2.79 (1H, m), 3.13 (1H, dd, J=4.6, 11.2 Hz), 3.43-3.70 (5H, m), 4.41, 4.87 (2H, AB-q, J=10.6 Hz), 4.46 (2H, s), 4.66, 4.71 (2H, AB-q, J=11.2 Hz), 4.81, 4.96 (2H, AB-q, J=10.6 Hz), 7.13-7.17 (2H, m), 7.24-7.38 (18H, m). FAB MS (positive); m/z 581 [M+H]⁺. High resolution FAB MS (positive); Calcd. for C₃₇H₄₅N₂O₄: 581.3373. Found: 581.3388.

N-(2-Isothiocyanatoethyl)-2,3,4,6-tetra-O-benzyl-1,5-dideoxy-1,5-imino-D-glucitol (15a). To a solution of the crude 14a (412 mg) obtained above in CH₂Cl₂ (15 ml) were added CS₂ (100 mg, 1.24 mmol) and Et₃N (202 mg, 2.00 mmol) at 24 °C under nitrogen. The solution was stirred for 1

h, and then to this solution were added Et₃N (150 mg) and 2-chloro-1-methylpyridinium iodide (260 mg, 1.02 mmol). This solution was stirred for 2.5 h at 24 °C under nitrogen, and concentrated *in vacuo* to give a mixture, which was diluted with EtOAc, washed with sat. NaHCO₃, H₂O, brine, dried over MgSO₄, and filtered. The filtrate was concentrated *in vacuo* to give a mixture, which was chromatographed on a silica gel column. Elution with cyclohexane-EtOAc (3:1) gave **15a** (156 mg, two steps from **13a**, 41%). IR v_{max} (film) 2170, 2110, 1648, 1590 cm⁻¹. 270 MHz ¹H NMR (CDCl₃) δ 2.34 (1H, t, J=10.8 Hz), 2.49 (1H, td, J=2.9, 9.0 Hz), 2.86 (1H, td, J=6.2, 12.0 Hz), 3.00-3.14 (2H, m), 3.38-3.52 (4H, m), 3.57-3.65 (3H, m), 4.44, 4.88 (2H, AB-q, J=10.9 Hz), 4.45 (2H, s), 4.65, 4.71 (2H, AB-q, J=11.6 Hz), 4.80, 4.95 (2H, AB-q, J=10.9 Hz), 7.13-7.19 (2H, m), 7.25-7.40 (18H, m). FAB MS (positive); m/z 609 [M+H]⁺. High resolution FAB MS (positive); Calcd. for C₃₇H₄₁N₂O₄S: 609.2830. Found: 609.2803.

N-[2-[[1R-(1 α ,2 β ,3 α ,4 β ,5 β)]-[1-(tert-Butyldimethylsilyloxymethyl)-1-hydroxy-2,3,4-tris(tert-butyldimethylsilyloxy)cyclopent-5-yl]aminothiocarbonyl]aminoethyl]-2,3,4,6-tetra-O-benzyl-1,5-dideoxy-1,5-imino-D-glucitol (16a). A solution of 15a (31 mg, 0.051 mmol) and 4 (36 mg, 0.057 mmol) in THF (0.5 ml) and Et3N (10 mg) was stirred at 20-25 °C for 2 days, concentrated in vacuo, and chromatographed on a silica gel plate. Development with cyclohexane-EtOAc (4:1) gave 16a (55 mg, 87%) as a gum. The Rf values of two starting materials (15a and 4) and the product (16a) were almost same. IR ν_{max} (film) 3335, 2955-2930, 1640 (shoulder) cm⁻¹. 270 MHz ¹H NMR (CDCl₃) δ 0.09-0.12 (24H, m), 0.90 (27H, broad s), 0.94 (9H, s), 1.53 (1H, m), 2.22 (1H, m), 2.43 (1H, m), 2.66 (1H, m), 3.00 (1H, m), 3.17 (1H, m), 3.43-3.99 (12H, m), 4.37-4.97 (8H, m), 7.09-7.12 (2H, m), 7.25-7.32 (18H, m). FAB MS (positive); m/z 1244 [M+H]+, 721. High resolution FAB MS (positive); Calcd. for C67H₁₁₀N₃O₉SSi₄: 1244.7046. Found: 1244.7064. Anal. Calcd. for C67H₁₀₉N₃O₉SSi₄ (1245.0): C, 64.64; H, 8.83; N, 3.38; S, 2.58. Found: C, 64.70; H, 8.48; N, 3.39; S, 2.63.

N-[3-[[1R-(1 α ,2 β ,3 α ,4 β ,5 β)]-[[1-(tert-Butyldimethylsilyloxymethyl)-1-hydroxy-2,3,4-tris(tert-butyldimethylsilyloxy)cyclopent-5-yl]aminothiocarbonyl]aminopropyl]-2,3,4,6-tetra-O-benzyl-1,5-dideoxy-1,5-imino-D-glucitol (16b). A solution of 14b (92 mg, 0.158 mmol) and 5 (107 mg, 0.158 mmol) in THF (0.20 ml) was warmed at 60-65 °C for 3 h with stirring, concentrated in vacuo, and chromatographed on a silica gel column. Elution with cyclohexane-EtOAc (4:1) gave 16b (166 mg, 83%) as a gum. IR v_{max} (film) 3100-2800, 2480 (w), 1640 cm⁻¹. 270 MHz ¹H NMR (CDCl₃) δ 0.09-0.12 (24H, m), 0.90-0.93 (36H, m), 1.26 (1H, broad), 1.50-1.80 (3H, m), 2.20 (1H, t, J=10.7 Hz), 2.32 (1H, t, J=9.2 Hz), 2.63 (1H, m), 2.80 (1H, m), 3.11 (1H, dd, J=4.7, 11.0 Hz), 3.33 (1H, m), 3.42-3.78 (8H, m), 3.90 (1H, m), 3.97 (1H, s), 4.35-4.97 (8H, m), 7.08-7.11 (2H, m), 7.22-7.38 (18H, m). FAB MS (positive); m/z 1260, 1259, 1258 [M+H]+, 738, 737, 735, 176. Anal. Calcd. for C₆₈H₁₁₁N₃O₉SSi₄ (1259.1): C, 64.87; H, 8.89; N, 3.34; S, 2.55. Found: C, 65.08; H, 8.93; N, 3.29; S, 2.59.

N-[2-[[\mathbb{R} -(1 α ,2 β ,3 α ,4 β ,5 β)]-(1-Hydroxymethyl-1,2,3,4-tetrahydroxycyclopent-5-yl)aminothiocarbonyl]aminoethyl]-2,3,4,6-tetra-O-benzyl-1,5-dideoxy-1,5-imino-D-glucitol (17a). To a solution of 16a (50 mg, 0.040 mmol) in MeOH (5 ml) was added a solution of 10% (w/w) HCl in MeOH (5 mL). The solution was stirred for 16 h at 24 °C, and concentrated *in vacuo* to give a hydrochloride of 17a (33 mg, quantitative) as a powder. IR ν_{max} (Nujpl) 3300 (broad), 1550 cm⁻¹. 270

MHz 1 H NMR (CDCl₃) δ 2.80-5.20 (26H, m), 6.87 (2H, bs), 7.07-7.40 (18H, m). FAB MS (positive); m/z 788 [M+H] $^{+}$. High resolution FAB MS (positive); Calcd. for C₄₃H₅₄N₃O₉S: 788.3579. Found: 788.3586.

N-[3-[[1R-(1 α ,2 β ,3 α ,4 β ,5 β)]-[1-Hydroxymethyl-1,2,3,4-(tetrahydroxy)cyclopent-5-yl]aminothiocarbonyl]aminopropyl]-2,3,4,6-tetra-O-benzyl-1,5-dideoxy-1,5-imino-D-glucitol Hydrochloride (17b). The similar treatment of 16b (473 mg) as described above gave 17b (314 mg, quantitative) as a powder. IR v_{max} (Nujpl) 3320 (broad), 1682, 1638 cm⁻¹. 270 MHz ¹H NMR (CDCl₃) δ 1.48-1.60 (2H, m), 2.09 (1H, m), 2.31 (1H, m), 2.46 (1H, m), 2.65-2.80 (2H, m), 3.08 (1H, m), 3.30 (1H, m), 3.40-3.71 (8H, m), 3.87-4.00 (3H, m), 4.32-4.90 (8H, m), 7.08-7.15 (2H, m), 7.21-7.33 (18H, m). FAB MS (positive); m/z 802 [M+H]+. High resolution FAB MS (positive); Calcd. for C₄₄H₅₆N₃O₉S: 802.3735. Found: 802.3748. *Anal.* Calcd. for C₄₄H₅₅N₃O₉S·HCl (838.5): C, 63.03; H, 6.73; N, 5.01; S, 3.82; Cl, 4.23. Found: C, 62.90; H, 6.76; N, 5.03; S, 3.67; Cl, 4.54.

 $N-[2-[[3aR-(3a\alpha,4\alpha,5\beta,6\alpha,6a\alpha)]-4-Hydroxymethyl-3a,5,6,6a-tetrahydro-4,5,6$ trihydroxy-4H-cyclopentoxazol-2-yl]aminoethyl]-2,3,4,6-tetra-O-benzyl-1,5-dideoxy-1,5-To a suspension of the hydrochloride 17a (56 mg, 0.068 mmol) in MeCN (5 imino-D-glucitol (18a). ml) was added 2-chloro-3-ethylbenzoxazolium tetrafluoroborate (42 mg, 0.156 mmol) at 0-5 °C under nitrogen with stirring. The suspension changed to a solution immediately. After stirring for 1 h at 0 °C, Et3N (170 mg, 1.68 mmol) was added. After stirring for 30 min at 0 °C, the reaction mixture was concentrated in vacuo, and chromatographed on a silica gel column. Elution with 10% MeOH in EtOAc, and MeOH-EtOAc (1:1) gave a mixture of silica gel and 18a, which was concentrated in vacuo, and diluted with EtOAc. The mixture was washed with sat. NaHCO3 and brine to remove the silica gel, dried over MgSO4, filtered, and concentrated in vacuo to give 18a (34 mg, 67%) as a yellowish powder. IR v_{max} (Nujpl) 3340 (broad), 1703, 1665 cm⁻¹. 270 MHz ¹H NMR (CDCl₃) δ 2.16 (1H, t, J=10.3 Hz), 2.32 (1H, m), 2.48 (1H, m), 2.87 (1H, m), 3.10 (1H, m), 3.20-3.30 (2H, m), 3.40-3.48 (2H, m), 3.50-3.67 (3H, m), 3.98 (2H, bs), 4.11 (1H, bs), 4.20 (1H, bs), 4.28-4.36 (2H, m), 4.44-5.09 (8H, m), 7.06-7.09 (2H, m), 7.16-7.40 (18H, m). FAB MS (positive); m/z 754 [M+H]⁺. High resolution FAB MS (positive); Calcd. for C₄₃H₅₂N₃O₉: 754.3705. Found: 754.3704.

N-[3-[[3aR-(3a α ,4 α ,5 β ,6 α ,6a α)]-4-Hydroxymethyl-3a,5,6,6a-tetrahydro-4,5,6-trihydroxy-4H-cyclopentoxazol-2-yl]aminopropyl]-2,3,4,6-tetra-O-benzyl-1,5-dideoxy-1,5-imino-D-glucitol (18b). The similar treatment of 17b (54 mg) as described above gave 18b (40 mg, 77%) as a yellowish powder. ^{1}H NMR (CDCl₃+DMF-d₇+D₂O) δ 3.62-4.83 (22H, m), 7.21-7.32 (20H, m). IR ν_{max} (Nujol) 3340 (broad), 1698 cm⁻¹. 270 MHz ^{1}H NMR (CDCl₃) δ 1.50-1.70 (2H, m), 2.04 (1H, broad), 2.25 (1H, m), 2.37 (1H, m), 2.70 (1H, m), 2.98-3.22 (3H, m), 3.37-3.70 (5H, m), 3.86-4.10 (3H, m), 4.20-4.95 (11H, m), 7.05-7.12 (2H, m), 7.20-7.37 (18H, m). FAB MS (positive) m/z 768 [M+H]+. High resolution FAB MS (positive), Calcd. for C44H54N3O9: 768.3858. Found: 768.3868. *Anal.* Calcd. for C44H53N3O9 (767.9): C, 68.82; H, 6.96; N, 5.47. Found: C, 68.30; H, 6.83; N, 5.48.

N-[2-[[3aR-(3aα,4α,5β,6α,6aα)]-4-Hydroxymethyl-3a,5,6,6a-tetrahydro-4,5,6-trihydroxy-4H-cyclopentoxazol-2-yl]aminoethyl]-1,5-dideoxy-1,5-imino-D-glucitol (19a).

A solution of 18a (34 mg, 0.045 mmol) in MeOH (10 ml) containing Pd(OH)₂ on carbon (50 mg, wet,

Degussa type, Pd content 20%) as a catalyst was stirred for 8 h at 60-65 °C under an atmosphere of hydrogen. The catalyst was filtered off, and the filtrate was concentrated *in vacuo*. The residue was chromatographed on Amberlite CG 50 (NH₄+ type/H⁺ type = 3/2, 12 ml). Elution with 0.5 M NH₃ aqueous solution (3 ml each), then 1.0 M NH₃ aqueous solution gave **19a** (13 mg, 72%) as a powder after lyophylization. IR v_{max} (Nujol) 3500-3100 (broad), 1705, 1660 cm⁻¹. 400 MHz ¹H NMR (D₂O) δ 2.13-2.22 (2H, m, CH₂-NH), 2.58 (1H, m, N-CH), 2.75 (1H, m, N-CH), 2.88 (1H, dd, J=4.9, 11.7 Hz, C1-H), 3.08 (1H, t, J=9.3 Hz), 3.10-3.20 (2H, m), 3.18 (1H, m), 3.36 (1H, m), 3.55, 3.65 (2H, AB-q, J=11.7 Hz, C6'-H₂), 3.67 (1H, d, J=2.4 Hz), 3.73-3.77 (2H, m), 3.99 (1H, dd, J=2.4, 4.4 Hz), 4.16 (1H, d, J=8.3 Hz), 4.75 (1H, dd, J=1.0, 8.8 Hz). ¹³C NMR (D₂O) δ 40.5 (CH₂-NH), 52.6 (CH₂-N), 58.0 (C1), 59.8 (C6 or C7'), 64.4 (C7' or C6), 71.0, 72.2, 75.8, 80.5, 82.3, 82.6, 85.2, 98.8 (C2, C3, C4, C5, C3a', C5', C6', C6a'), 163.7 (C2'). FAB MS (positive); m/z 394 [M+H]+. High resolution FAB MS (positive); m/z Calcd. for C₁₅H₂₈N₃O₉: 394.1825. Found: 394.1834.

 $N-[3-[[3aR-(3a\alpha,4\alpha,5\beta,6\alpha,6a\alpha)]-4-Hydroxymethyl-3a,5,6,6a-tetrahydro-4,5,6-$

trihydroxy-4*H*-cyclopentoxazol-2-yl]aminopropyl]-1,5-dideoxy-1,5-imino-D-glucitol (19b). A solution of 18b (50 mg, 0.065 mmol) in MeOH (10 ml) containing Pd(OH)₂ on carbon (55 mg, wet, Degussa type, Pd content 20%) as a catalyst was hydrogenolized for 24 h at 50-55 °C. The reaction mixture was treated as described above to give 19b (26 mg, 98%) as a powder after lyophylization. [α]D²⁴ +56.0° (c 0.6, H₂O). IR ν_{max} (KBr) 3340 (broad), 2926, 2337 (w), 1706, 1659 cm⁻¹. 400 MHz ¹H NMR (D₂O) δ 1.50-1.57 (2H, m, C-CH₂-C), 2.03-2.12 (2H, m, C5H-N, N-C1H^{β}), 2.47 (1H, m, N-CHC), 2.59 (1H, m, N-CHC), 2.80 (1H, dd, *J*=5.1, 11.4 Hz, N-C1H^{α}), 2.98-3.08 (3H, m, N-CH₂C, C3H), 3.17 (1H, t, *J*=9.5 Hz, C4H), 3.35 (1H, m, C2H), 3.53-3.71 (4H, m, C6H₂OH, C4'CH₂OH), 3.78 (1H, d, *J*=4.4 Hz, C5'H), 4.03 (1H, dd, *J*=2.4, 4.4 Hz, C6'H), 4.20 (1H, d, *J*=8.3 Hz, C3a'H), 4.86 (1H, d, *J*=8.3 Hz, C6a'H). 400 MHz ¹³C NMR (D₂O) δ 25.2, 42.8, 51.4, 57.5, 59.7, 64.0, 67.3, 71.1, 72.2, 73.5, 80.5, 82.1, 82.3, 85.2, 90.9, 163.7. FAB MS (positive); m/z 408 [M+H]⁺. High resolution FAB MS (positive); m/z Calcd. for C₁₆H₃₀N₃O₉: 408.2027. Found: 408.1952.

N-(3-Aminopropyl)-1,5-dideoxy-1,5-imino-D-glucitol (20). Compound 14b (95 mg) in MeOH (15 ml) was hydrogenolyzed 50-55 °C for 16 h using Pd(OH)₂ on carbon (105 mg). The soltion was filtered, and concentrated *in vacuo* to give 20 (31 mg, 86%), which was dissolved in H₂O, and lyophilized. IR v_{max} (KBr) 3352 (broad) cm⁻¹. 400 MHz ¹H NMR (D₂O) δ 1.54 (2H, quintet, J=7.2-7.6 Hz), 2.05 (1H, m), 2.07 (1H, t, J=11.4 Hz), 2.44 (1H, m), 2.58-2.69 (3H, m), 2.85 (1H, dd, J=4.9, 11.4 Hz), 3.07 (1H, t, J=9.4 Hz), 3.18 (1H, t, J=9.4 Hz), 3.35 (1H, m), 3.65 (1H, dd, J=2.7, 12.8 Hz), 3.72 (1H, dd, J=2.3, 12.8 Hz). 400 MHz ¹³C NMR (D₂O) δ 25.71 (H₂C), 40.94 (H₂C), 51.40 (H₂C), 57.42 (H₂C), 59.58 (H₂C), 67.44 (HC), 71.05 (HC), 72.24 (HC), 80.58 (HC). FAB MS (positive); m/z 221 [M+H]⁺. High resolution FAB MS (positive); m/z Calcd. for C9H₂₁N₂O₄: 221.1501. Found: 221.1504.

 $[1R-(1\alpha,2\beta,3\alpha,4\beta,5\beta)]$ -5-[(Butylaminothiocarbonyl)amino]-1-(tert-

butyldimethylsilyloxymethyl)-2,3,4-tris(tert-butyldimethylsilyloxy)cyclopentane-1-ol (21). A solution of amine 4 (64 mg, 0.10 mmol), butyl isothiocyanate (20 mg, 0.17 mmol) and Et3N (10 mg,

0.10 mmol) in THF (0.5 ml) was allowed to stand at room tempetature for 16 h. Reaction mixture was

concentrated *in vacuo*, and chromatographed on a silica gel column. Elution with cyclohexane-EtOAc (9:1) gave 21 (55 mg, 73%) as a solid, mp 141-142 °C (from EtOAc-hexane). IR v_{max} (Nujol) 3335, 1648 cm⁻¹. 270 MHz ¹H NMR (CDCl₃) δ 0.07-0.15 (24H, m), 0.85-0.96 (39H, m), 1.31-1.41 (2H, m), 1.51-1.62 (2H, m), 3.24-3.53 (2H, m), 3.68-4.20 (6H, m). FAB MS (positive); m/z 753, 752, 751 [M+H]⁺. High resolution FAB MS (positive); m/z Calcd. for C₃₅H₇₈N₂O₅SSi₄: 751.4797. Found: 751.4756.

[1R-(1 α ,2 β ,3 α ,4 β ,5 β)]-5-[(Butylaminothiocarbonyl)amino]-1-hydroxymethyl-cyclopentane-1,2,3,4-tetraol (22). A solution of amine 21 (44 mg, 0.06 mmol) in MeOH (4 mL) and 10% HCl in MeOH (4 ml, w/w) was stirred at 24 °C for 4 h. Reaction mixture was concentrated *in vacuo* to give 22 (17 mg, quantitatively) as a gum, which was employed for the next reaction without purification. IR ν_{max} (film) 3320, 2959, 2933, 2880, 1620, 1563, 1555 cm⁻¹. 270 MHz ¹H NMR (CD₃COCD₃-D₂O) δ 0.92 (3H, t, J=7.3 Hz), 1.30-1.42 (2H, m), 1.52-1.63 (2H, m), 3.38-3.64 (3H, m), 3.82 (1H, bs), 3.92 (1H, d, J=7.0 Hz), 3.97 (1H, m), 3.99, 4.14 (2H, AB-q, J=9.1 Hz). FAB MS (positive); m/z 295 [M+H]⁺. High resolution FAB MS (positive); m/z Calcd. for C₁₁H₂₂N₂O₅S: 295.1384. Found: 295.1304.

 $[3aR-(3a\alpha,4\alpha,5\beta,6\alpha,6a\alpha)]-2$ -Butylamino-4-hydroxymethyl-4,5,6,6a-tetrahydro-3aHcyclopentaoxazole-4,5,6-triol (23). To a solution of thiourea 22 (17 mg, 0.06 mmol) in MeCN (2 ml) was added 2-chloro-3-ethylbenzoxazolium tetrafluoroborate (29 mg, 0.11 mmol) at 0 °C with stirring under nitrogen atmosphere. After 1 h stirring at 0 °C, Et₃N (50 mg, 0.50 mmol) was added to the mixture. After stirring at 0 °C for 30 min, reaction mixture was concentrated in vacuo and was then dissolved in H2O (2 ml). The aqueous solution was washed with EtOAc (l ml). The aqueous layer was charged on an Amberlite CG 50 $(NH4^+ type/H^+ type = 3/2, 15 ml)$ column for purification. Elution with 0.5 M aqueous NH4OH (4 ml x 10) and then 1.5 M aqueous NH4OH (4 ml each) gave 23 (7.0 mg, two steps 46%) on the No. 14 as a powder after lyophylization. IR v_{max} (film) 3375, 2960, 2930, 2870, 1659 cm⁻¹. 400 MHz 1 H NMR (CDCl₃) δ 0.70 (3H, t, J=7.3 Hz), 1.09-1.18 (2H, m), 1.25-1.36 (2H, m), 2.87-2.99 (2H, m, CH₂N), 3.52, 3.63 (2H, AB-q, J=12.0 Hz), 3.74 (1H, d, J=4.9 Hz, C5-H), 3.95 (1H, dd, J=2.5, 4.9 Hz, C6-H), 4.11 (1H, d, J=8.8Hz, C_{3a} -H), 4.66 (1H, m, C_{6a} -H). 400 MHz ¹³C NMR (D₂O) δ 15.3 (CH₃), 21.5 (CH₂), 33.1 (CH₂), 44.2 (CH2-NH), 64.6 (CH2OH), 76.3 (C3a), 82.3, 82.4, 88.9 (C5, C6, C6a), 85.0 (C4), 164.1 (C2). FAB MS (positive); m/z 261 [M+H]⁺. High resolution FAB MS (positive); m/z Calcd. for C₁₁H₂₁N₂O₅S: 261.1451. Found: 261.1477.

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